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Recent Events Dominate Interdomain Lateral Gene Transfers Between Prokaryotes and Eukaryotes and, with the Exception of Endosymbiotic Gene Transfers, Few Ancient Transfer Events Persist

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Electronic supplementary material is available at http://dx.doi.org/10.1098/rstb.2014.0324 or via http://rstb.royalsocietypublishing.org. Recent events dominate interdomain lateral gene transfers between prokaryotes and eukaryotes and, with the exception of endosymbiotic gene transfers, few ancient transfer events persist

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While there is compelling evidence for the impact of endosymbiotic gene transfer (EGT; transfer from either mitochondrion or chloroplast to the nucleus) on genome evolution in eukaryotes, the role of interdomain transfer from bacteria and/or archaea (i.e. prokaryotes) is less clear. Lateral gene transfers (LGTs) have been argued to be potential sources of phylogenetic information, particularly for reconstructing deep nodes that are difficult to recover with traditional phylogenetic methods. We sought to identify interdomain LGTs by using a phylogenomic pipeline that generated 13 465 single gene trees and included up to 487 eukaryotes, 303 bacteria and 118 archaea. Our goals include searching for LGTs that unite major eukaryotic clades, and describing the relative contributions of LGT and EGT across the eukaryotic tree of life. Given the difficulties in interpreting single gene trees that aim to capture the approximately 1.8 billion years of eukaryotic evolution, we focus on presence-absence data to identify interdomain transfer events. Specifically, we identify 1138 genes found only in prokaryotes and representatives of three or fewer major clades of eukaryotes (e.g. Amoebozoa, Archaeplastida, Excavata, Opisthokonta, SAR and orphan lineages). The majority of these genes have phylogenetic patterns that are consistent with recent interdomain LGTs and, with the notable exception of EGTs involving photosynthetic eukaryotes, we detect few ancient interdomain LGTs. These analyses suggest that LGTs have probably occurred throughout the history of eukaryotes, but that ancient events are not maintained unless they are associated with endosymbiotic gene transfer among photosynthetic lineages.

1. Inferences about lateral and endosymbiotic gene transfer

The impact of lateral gene transfer (LGT) is best known in bacteria where the phenomenon of the rapid spread of antibiotic resistance among bacterial strains/species highlights the importance of this process in our own lives [1,2]. Analyses of first single genes and more recently whole genomes have demonstrated large numbers of LGTs among bacteria and archaea [3–6] and have contributed to discussions on the nature of species in these clades [7–9]. Less clear is the role of LGT in the evolution of eukaryotes, which may result from the use of the animals as models for evolutionary principles in eukaryotes given that the sequestration of the germline in triploblastic animals probably created a barrier to LGT [10–14]. There is a growing literature on LGTs involving eukaryotes, including microbial lineages [4,5,13], fungi [15,16], plants [17,18] and some animal lineages [19,20]. The bulk of these analyses focus on what might be termed 'tip down' approaches, asking about the impact of LGTs within clades rather than across the eukaryotic tree of life.

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In contrast to the debate on the role of LGT in eukaryotes, the past few decades have seen a substantial rise in descriptions of cases of endosymbiotic gene transfer (EGT): gene transfer from mitochondrion or plastid to the nucleus [21–24]. For example, roughly 15–20% of plant genomes are probably derived from plastid genes [25,26], and somewhere between 10% and 50% of the mitochondrial proteome is derived from nuclear encoded genes of alpha-proteobacterial origin [27,28]. Hence, transfer of genes from organelle to nucleus within a lineage is now well established [22,23,29,30].

Arguments have been made that LGTs can be used as evidence for ancient relationships, and that such data can be useful in reconstructing ancient relations [31,32]. For example, several authors have argued that there was a pulse of LGTs from various bacteria, including Chlamydiae, to the last common ancestor of Archaeplastida [33,34]. Analyses of networks generated by shared LGTs can be informative in discerning shared history among bacteria and/or archaea (hereafter termed prokaryotes) [35,36]. Abby *et al.* [35] use the distribution of LGT events in reconstructing relationships among bacteria and archaea, whereas Szollosi *et al.* [36] demonstrate the power of using LGTs to reconstruct the pattern and timing of events within bacterial genera and species.

Inferring ancient LGTs can be very difficult [22,37], which is why we focus on presence-absence data consistent with interdomain LGT. Inferring the transfer of single genes among divergent lineages of eukaryotes based on the topology of single gene trees is challenging given that we would be asking approximately 200-300 amino acids to estimate events as old as approximately 1.8 billion years (an estimate of the timing of eukaryotic origins [38,39]). In addition, errors in phylogenetic reconstruction such as long branch attraction and incomplete taxon sampling can mislead interpretations of lateral events based on tree topologies [22,40,41]. Perhaps most importantly for the analyses presented here, the prevalence of gene loss over evolutionary time [42] confounds interpretation of lateral events as genes present in bacteria plus only a few non-sister eukaryotic lineages may have been lost in other eukaryotic lineages. In other words, parallel gene loss among disparate lineages can mistakenly be interpreted as LGT among lineages retaining any given gene.

2. Estimating interdomain lateral gene transfers across the eukaryotic tree of life

Mindful of the perils and pitfalls of interpreting ancient gene transfer events, we set out to assess the tempo of interdomain LGTs and EGTs in the early evolution of eukaryotes by focusing on presence-absence data. Such analyses are possible because of our development of a phylogenomic pipeline that focuses on inclusion of a broad diversity of microbial lineages [43,44]. In brief, we start with clusters of homologous sequences (i.e. genes) as determined in OrthoMCL [45,46] and then add diverse taxa to end up with a sample of up to 487 eukaryotes, 303 bacteria and 118 archaea. The pipeline uses custom PYTHON scripts and third-party tools such as NEEDLE [47] to remove sequences that are either too similar (e.g. alleles) or too divergent (e.g. poor-quality transcripts, sequences sharing only motifs). Multi-sequence alignments are generated and refined using GUIDANCE [48] and single gene trees are constructed using RAXML [49,50] using parameters from Grant & Katz [44,51]. We then used custom scripts to identify orthologues present in at least 10 taxa, at least three of which are bacteria/archaea, and a monophyletic clade of two or more eukaryotic sequences. Because of our focus on interdomain LGTs, the specifics of the resulting tree topologies (i.e. relationships among eukaryotes or among prokaryotes) are not critical, though we did require that eukaryotic sequences form a monophyletic clade.

Analyses of 13 465 genes, which included up to 908 diverse lineages (table 1 and electronic supplementary material, table S1), yielded 1138 genes that met our criteria for possible examples of interdomain transfers between prokaryotes and eukaryotes (table 2). We identified interdomain gene transfer events based on the presence of genes in prokaryotes plus members of three or fewer major eukaryotic clades (e.g. Opisthokonta, Archaeplastida; table 1) that formed a monophyletic group in the RAXML trees automatically generated by the pipeline [44]. This distribution was chosen because we think it is likely that orthologues present in four or more of the major eukaryotic clades were probably present in the last eukaryotic common ancestor (LECA).

We then categorized the 1138 genes based on their distributions in major (MC) and minor (mc) eukaryotic clades. Under this notation, genes in the 1MC1mc category are found in prokaryotes plus only one minor clade in only one major clade of eukaryotes, whereas genes in 3MC2mc are found in three major clades of eukaryotes with at least two minor clades in one of these major clades (table 2; electronic supplementary material, table S2). Because we controlled the names of our taxa to reflect major and minor clades (table 1; electronic supplementary material, table S1; [43,44]), categorizing genes was readily accomplished using the P4 package (https://code. google.com/p/p4-phylogenetics/). We also inspected the resulting trees to determine if a single or monophyletic minor clade of bacteria or archaea were sister to the eukaryotic sequences (electronic supplementary material, table S2), though we recognize the caution needed in interpreting these relationships given the likelihood of prokaryote-prokaryote LGT transfer [52].

3. The majority of putative interdomain lateral gene transfers appear to be recent events

Inspection of the 1138 genes that match our criteria for putative interdomain LGT reveal a striking pattern as over half of the putative interdomain LGTs (606 of 1138) involve only one minor clade nested within one major clade of eukaryotes (e.g. metazoa (Op_me) or ciliates (Sr_ci); table 2 and electronic supplementary material, S2). In fact, the greatest number of interdomain LGTs in this category are found in only one minor clade within the Opisthokonta (290 genes), Archaeplastida (170 genes) and then SAR (Stramenopila + Alveolata + Rhizaria; 59 genes; table 3). To exemplify this pattern, we include an example of one of the resulting trees for a putative carboxymuconolactone decarboxylase enzyme (OG5_141348 from OrthoMCL), which is found only in bacteria, archaea and animals (figure 1). The large number of interdomain LGTs into fungi (196 inferred; figure 2; electronic supplementary material, table S1) is consistent with numerous studies [15,53], but a broader comparison of our data with published cases of putative LGT is not easily done as we used more restrictive criteria (i.e. eukaryotes must be monophyletic) than most.

Table 1. Taxon sampling and major_minor clade abbreviations used in the analyses for the 487 eukaryotes, 303 bacteria and 118 archaea. Individual species/ strain names are found in the electronic supplementary material, table S1. *n*, number of species/strains included in each category. Naming system is based largely on NCBI taxonomy, though no assumption is made on equivalency of rank for major (first abbreviation) and minor (second abbreviation) clades. The five major clades of eukaryotes each have a unique code (Op, Opisthokonta; Am, Amoebozoa; Ex, Excavata; PI, Archaeplastida (Plantae); Sr, SAR (Stramenopila + Alveolata + Rhizaria)) and we use the abbreviation EE (everything else) to capture the non-monophyletic 'orphan' lineages (table 1).

Am_acAmoebozoa: Acanthamoebidae1Ar_crarchaea: Crenarchaeota27Am_arAmoebozoa: Archamoebae5Ar_earchaea: Euryarchaeota77Am_daAmoebozoa: Discosea1Ar_koarchaea: Korarchaeota1Am_diAmoebozoa: Dictyostellida3Ar_naarchaea: Nanoarchaeota1Am_fiAmoebozoa: Filamoeba1Ar_nharchaea: Nanohaloarchaeota3Am_fiAmoebozoa: incertae sedis4Ar_paarchaea: Parvarchaeota2Am_myAmoebozoa: Mycetozoa2Ar_tharchaea: Thaumarchaeota3Am_vaAmoebozoa: Vannellidae2Ba_acbacteria: Actinobacteria31EE_aporphan: Apusozoa2Ba_aqbacteria: Aquificae5
Am_arAmoebozoa: Archamoebae5Ar_earchaea: Euryarchaeota77Am_daAmoebozoa: Discosea1Ar_koarchaea: Korarchaeota1Am_diAmoebozoa: Dictyostellida3Ar_naarchaea: Nanoarchaeota1Am_fiAmoebozoa: Filamoeba1Ar_nharchaea: Nanohaloarchaeota3Am_fiAmoebozoa: incertae sedis4Ar_paarchaea: Parvarchaeota2Am_myAmoebozoa: Mycetozoa2Ar_tharchaea: Thaumarchaeota7Am_vaAmoebozoa: Vannellidae2Ba_acbacteria: Actinobacteria31EE_aporphan: Apusozoa1Ba_aqbacteria: Aquificae5
Am_daAmoebozoa: Discosea1Ar_koarchaea: Korarchaeota1Am_diAmoebozoa: Dictyostellida3Ar_naarchaea: Nanoarchaeota1Am_fiAmoebozoa: Filamoeba1Ar_nharchaea: Nanohaloarchaeota3Am_isAmoebozoa: incertae sedis4Ar_paarchaea: Parvarchaeota2Am_myAmoebozoa: Mycetozoa2Ar_tharchaea: Thaumarchaeota7Am_vaAmoebozoa: Vannellidae2Ba_acbacteria: Actinobacteria31EE_aporphan: Apusozoa1Ba_aqbacteria: Aquificae5
Am_diAmoebozoa: Dictyostellida3Ar_naarchaea: Nanoarchaeota1Am_fiAmoebozoa: Filamoeba1Ar_nharchaea: Nanohaloarchaeota3Am_isAmoebozoa: incertae sedis4Ar_paarchaea: Parvarchaeota2Am_myAmoebozoa: Mycetozoa2Ar_tharchaea: Thaumarchaeota7Am_vaAmoebozoa: Vannellidae2Ba_acbacteria: Actinobacteria31EE_aporphan: Apusozoa1Ba_adbacteria: Acidobacteria1EE_brorphan: Breviatea2Ba_aqbacteria: Aquificae5
Am_fiAmoebozoa: Filamoeba1Ar_nharchaea: Nanohaloarchaeota3Am_isAmoebozoa: incertae sedis4Ar_paarchaea: Parvarchaeota2Am_myAmoebozoa: Mycetozoa2Ar_tharchaea: Thaumarchaeota7Am_vaAmoebozoa: Vannellidae2Ba_acbacteria: Actinobacteria31EE_aporphan: Apusozoa1Ba_adbacteria: Acidobacteria1EE_brorphan: Breviatea2Ba_aqbacteria: Aquificae5
Am_isAmoebozoa: incertae sedis4Ar_paarchaea: Parvarchaeota2Am_myAmoebozoa: Mycetozoa2Ar_tharchaea: Thaumarchaeota7Am_vaAmoebozoa: Vannellidae2Ba_acbacteria: Actinobacteria31EE_aporphan: Apusozoa1Ba_adbacteria: Acidobacteria1EE_brorphan: Breviatea2Ba_aqbacteria: Aquificae5
Am_myAmoebozoa: Mycetozoa2Ar_tharchaea: Thaumarchaeota7Am_vaAmoebozoa: Vannellidae2Ba_acbacteria: Actinobacteria31EE_aporphan: Apusozoa1Ba_adbacteria: Acidobacteria1EE_brorphan: Breviatea2Ba_aqbacteria: Aquificae5
Am_vaAmoebozoa: Vannellidae2Ba_acbacteria: Actinobacteria31EE_aporphan: Apusozoa1Ba_adbacteria: Acidobacteria1EE_brorphan: Breviatea2Ba_aqbacteria: Aquificae5
EE_aporphan: Apusozoa1Ba_adbacteria: Acidobacteria1EE_brorphan: Breviatea2Ba_aqbacteria: Aquificae5
EE_brorphan: Breviatea2Ba_aqbacteria: Aquificae5
EE_cr orphan: Cryptophyta 13 Ba_ar bacteria: Armatimonadetes 1
EE_haorphan: Haptophyceae16Ba_babacteria: Bacteroidetes21
EE_isorphan: incertae sedis6Ba_bcbacteria: Chlorobi3
EE_ka orphan: Katablepharidophyta 1 Ba_bi bacteria: Ignavibacteriae 2
Ex_euExcavata: Euglenozoa23Ba_cabacteria: Caldiserica1
Ex_fo Excavata: Fornicata 6 Ba_cd bacteria: Chlamydiae 6
Ex_heExcavata: Heterolobosea4Ba_chbacteria: Chloroflexi9
Ex_isExcavata: incertae sedis1Ba_crbacteria: Chrysiogenetes1
Ex_ja Excavata: Jakobida 5 Ba_cv bacteria: Verrucomicrobia 4
Ex_maExcavata: Malawimonadidae2Ba_cybacteria: Cyanobacteria41
Ex_oxExcavata: Oxymonadida1Ba_debacteria: Deinococci7
Ex_paExcavata: Parabasalia4Ba_dfbacteria: Deferribacteres2
Op_ch Opisthokonta: Choanoflagellida 5 Ba_di bacteria: Dictyoglomi 2
Op_fu Opisthokonta: fungi 40 Ba_el bacteria: Elusimicrobia 1
Op_icOpisthokonta: Ichthyosporea3Ba_fbbacteria: Firmicute, Bacilli17
Op_isOpisthokonta: incertae sedis1Ba_fcbacteria: Firmicute, Clostridia13
Op_me Opisthokonta: Metazoa 61 Ba_fn bacteria: Firmicute, Negativicutes 2
Pl_glArchaeplastida: Glaucocystophytes3Ba_fubacteria: Fusobacteria5
Pl_grArchaeplastida: green algae61Ba_gebacteria: Gemmatimonadetes1
Pl_rh Archaeplastida: Rhodophyta 20 Ba_is bacteria: incertae sedis 2
Sr_ap SAR: Apicomplexa 18 Ba_me bacteria: Melainabacteria 1
Sr_ch SAR: Chromerida 2 Ba_ni bacteria: Nitrospirae 3
Sr_ciSAR: Ciliophora27Ba_pabacteria: Alphaproteobacteria24
Sr_diSAR: Dinophyceae33Ba_pbbacteria: Betaproteobacteria17
Sr_is SAR: incertae sedis 1 Ba_pd bacteria: Deltaproteobacteria 14
Sr_peSAR: Perkinsida2Ba_pgBacteria: Gammaproteobacteria31
Sr_rhSAR: Rhizaria22Ba_plbacteria: Planctomycetes6
Sr_stSAR: Stramenopila84Ba_spbacteria: Spirochaetes9
Ba_sy bacteria: Synergistetes 4
Ba_te bacteria: Tenericutes 7
Ba_th bacteria: Thermotogae 7
Ba_ts bacteria: Thermodesulfobacterium 2

Table 2. Number of genes (orthologous groups) analysed for this study by category, organized based on those found in one, two or three major clades (MC) and noting the number of minor clades (mc). Using a starting set of 13 465 alignments/trees, we selected genes that met our criteria of having at least 10 sequences, at least three of which are bacteria/archaea, and a monophyletic clade of eukaryotes based on output of phylogenomic pipeline [43,44]. #MC refers to the number of major clades, including the non-monophyletic orphans (table 1). #mc refers to number of minor clades: 1mc = only one minor clade; $2mc \ge two minor clades$ in at least one major clade. Individual genes are listed in electronic supplementary material, table S2.

abbreviation	description	n
1MC1mc	involving only one major clade and one minor clade	606
1MC2mc	involving only one major clade and at least two minor clades	77
2MC1mc	involving two major clades and only one minor clade in each	250
2MC2mc	involving two major clades and at least two minor clades in one major clade	140
3MC1mc	involving three major clades and only one minor clade in each	15
3MC2mc	involving three major clades and at least two minor clades in one major clade	50
total number		1138

Table 3. Recent interdomain LGTs involving prokaryotes and a single major clade of eukaryotes. Minor clade refers to nested taxa within the five major eukaryotic clades such as fungi with Opisthokonta or Apicomplexa within SAR. Numbers in parentheses are the number of species sampled (see electronic supplementary material, table S1). Individual genes are listed in electronic supplementary material, table S2.

	Amoebozoa (24)	orphans (37)	Excavata (49)	Opisthokonta (111)	Archaeplastida (85)	SAR (195)
one minor clade	42	2	43	290	170	59
\geq two minor clades	3	0	3	31	29	11
total	45	2	46	321	199	70

Multiple factors probably impact these patterns including the number of lineages sampled within major clades (111, 85 and 195 for Opisthokonta, Archaeplastida and SAR, respectively; table 3) and the nature of the data used in our pipeline (e.g. genome sequence versus RNAseq [43,44]). Also, many lineages within Opisthokonta and Archaeplastida have relatively large genomes (e.g. http://data.kew.org/cvalues/, http://www.genomesize.com/) which may increase the probability of retention of ancient LGTs. Interdomain LGT events seem to be underrepresented among lineages within the Excavata, even though the pipeline includes whole genome data from several genomes in this clade (e.g. the genera Leishmania, Trypanosoma, Giardia, Trichomonas). As Excavata with whole-genome sequences are nearly all parasites and may have experienced considerable gene loss, analyses of free-living Excavata are likely to reveal additional examples of interdomain LGTs in this major eukaryotic clade.

We inspected the five LGTs that appear to define major clades (see asterisk symbol in figure 2) and, given the caveats discussed in the following, propose these be considered as only candidate synapomorphies until additional diverse lineages of eukaryotes are sampled. For example, the one gene that is found in at least three lineages of Opisthokonta (OG5_146700) is a hypothetic protein present in our pipeline in a subset of archaea, fungi, choanoflagellates and only one metazoan (electronic supplementary material, table S2). The three genes that may serve as synapomorphies for the SAR clade are patchily distributed (electronic supplementary material, table S2) and have diverse functions: a putative pyruvoyl tetrahydropterin synthase (OG5_141276), a penicillin

amidase family protein (OG5_136942) and a putative deoxyribodipyrimidine photolyase (OG5_168036). Perhaps most optimistic as a synapomorphy is the one LGT at the base of Excavata, an acyl-CoA synthetase (OG5_146682), which has a relatively broad distribution given our sampling; it is found in Fornicata (*Giardia* and *Spironucleus*), Parabasalia (*Trichomonas*), Heterolobosea (*Sawyeria*) and Euglenozoa (*Euglena*).

4. The bulk of putative ancient interdomain gene transfers are likely to be endosymbiotic gene transfers

In contrast to the many recent interdomain LGTs, we see no compelling evidence for a pulse of LGT events that occurred in the common ancestors of major eukaryotic clades with the exception of gene transfers shared among clades with many photosynthetic members (table 4; electronic supplementary material, table S2; figure 2). The greatest numbers of putative interdomain LGTs involve clades with predominantly photosynthetic lineages (e.g. Archaeplastida (e.g. red and green algae), SAR (e.g. dinoflagellates, stramenopiles) and the 'orphan' Cryptophyta and Haptophyta (table 4 and electronic supplementary material, table S2). For example, there are 106 genes present in prokaryotes plus Archaeplastida plus SAR (table 4). Of the total of 455 genes that unite either two or three major clades of eukaryotes, 64 have gene tree topologies where photosynthetic eukaryotes are sister to cyanobacteria (electronic supplementary material, table S2). Retaining



Figure 1. An example of a recent interdomain LGT from prokaryotes to one minor dade (Metazoa) in one major clade (Opisthokonta) of eukaryotes. This tree exemplifies the many recent (e.g. 1MC1mc) interdomain transfers detected in this study (table 2 and figure 3). Abbreviations of taxa are as in table 1 and electronic supplementary material, S1, and the number following each name is a unique identifier from either OrthoMCL or GenBank. Analyses of this gene used PROTGAMMA, the best-fitting LG model and default parameters as implemented in RaxML [49,50]. Most nodes are poorly supported and only bootstrap values above 80% are shown. Monophyletic clades are marked with solid lines, whereas the complex relationships among prokaryotes in the dashed clades probably represent a combination of poorly resolved phylogeny, LGT among prokaryotes and gene loss.

cyanobacterial sisters is neither a strict requirement nor predictor of EGT as subsequent LGTs among prokaryotes in the approximately 1 billion years since the acquisition of plastids may confound the signature of EGT [38]; nevertheless, these 64 gene trees provide additional support for EGT.

While there are numerous other genes that unite two (tables 3 and 4) or three (electronic supplementary material, table S2) major clades, there are no clear patterns except that the highest numbers of putative events are among clades with many species sampled and/or lineages with large genomes (table 3 and electronic supplementary material, table S2). In other words, interdomain LGTs do not appear to be a good source for synapomorphies for deep eukaryotic relationships. With the exception of events that unite photosynthetic lineages, we suspect that the putative LGT events counted in table 4 are either interdomain LGTs followed by loss or a combination of interdomain and intradomain LGTs. For example, a gene found only in bacteria plus two major clades of eukaryotes could be the result of (i) vertical ancestry plus loss in the remaining major clades or (ii) interdomain LGT followed by intradomain LGT. Discerning between such hypotheses is very challenging given the limited power within single gene trees.

To assess whether shared gene transfer events unite major clades of eukaryotes, we also used the software Coevolution Of Presence–Absence Patterns (CoPAP) [54]. COPAP is designed to detect patterns of co-evolving genes in presence–absence data from diverse lineages and uses efficient probabilistic models to assess the significance of relationships [54]. We inverted our data to ask whether there are taxa that share significant numbers of LGT events from putative cases of interdomain gene transfers. We used a *p*-value of 0.05 as cut-off for interactions, a star phylogeny for relationships among the 455 genes that were found in two or three major eukaryotic clades (2MC or 3MC; electronic supplementary material, table S2), plus default parameters as implemented by COPAP (http://copap.tau.ac.il/).

Only a small number of significant interactions are supported from analyses of presence–absence of 456 genes shared among two or three major clades of eukaryotes (figure 3). One significant network contains the predominantly photosynthetic lineages of dinoflagellates (Sr_di), glaucophytes (Pl_gl), red algae (Pl_rh), cryptophytes (EE_cr) and haptophytes (EE_ha). A second network contains the predominantly photosynthetic stramenopiles (Sr_st) plus green algae (Pl_gr), though this network is disconnected from the other photosynthetic lineages (figure 3). Uniting photosynthetic lineages is consistent with EGT from plastid to nucleus. Such transfers involve members of the Archaeplastida, the lineage descended from an ancestor that had a primary acquisition of plastids from cyanobacteria [58], plus the remaining lineages of photosynthetic eukaryotes (e.g. diatoms, brown algae, cryptophytes,



*= found in at least three minor clades

Figure 2. Recent LGT events mapped onto representative lineages from the eukaryotic tree of life. Numbers at nodes represent the LGT events in table 3, and the synthetic tree is arbitrarily rooted on Opisthokonta. Numbers marked by asterisk are found in at least three minor clades within major clades and may represent synapomorphies for major clades. Arrows on the right mark shared putative LGTs found between non-sister minor clades. Numbers in green (grey) in parentheses are genes where eukaryotes fall sister to cyanobacteria and are hence putative EGTs. For simplicity, only a subset of lineages are included here and full taxonomic distributions can be found in the electronic supplementary material, tables S1 and S2. (Online version in colour.)

Table 4. Putative interdomain transfers shared between two major eukaryotic clades. Parentheses contain the number of putative interdomain LGTs present in only one minor clade of both major clades, which suggests these may not be shared ancient events. Individual genes are listed in electronic supplementary material table S2.

	orphans	Excavata	Opisthokonta	Archaeplastida	SAR
Amoebozoa	1 (1)	11 (11)	28 (18)	9 (8)	8 (5)
orphans		0	7 (1)	17 (13)	12 (6)
Excavata		—	26 (21)	8 (7)	9 (4)
Opisthokonta			—	71 (58)	76 (41)
Archaeplastida				—	106 (55)
SAR					

haptophytes and dinoflagellates) that acquired photosynthesis through secondary endosymbiosis [59,60].

The only other significant cluster detected by COPAP is the pairing of the archamoeba parasites *Entamoeba* spp. (Am_ar) with parabasalids including *Trichomonas* (Ex:pa; figure 3). An association between *Entamoeba* spp. and parabasalids was first described by several authors [55-57] before we found support of this hypotheses through analyses of interdomain and intradomain LGT involving *Entamoeba* spp. [51]. Given that we are using the same dataset here,

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Figure 3. Significant networks among lineages as determined by COPAP [54] based on presence – absence of LGTs. The green (dark) taxa are predominantly photosynthetic, and networks involving these minor clades indicate the potential influence of EGT on photosynthetic lineages. There is also a significant relationship between *Entamoeba* spp. (Am_ar) and parabasalids (Ex_pa) as has been previously observed [51,55–57]. The linking of fungi (Op_fu) and microbial opisthokonts (Op_ot; other Opisthokonta = lchthyosporea plus lineages that are incertae sedis) probably represents shared retention of LGT events. (Online vesion in colour.)

our finding this association is not surprising, though COPAP evaluates patterns of gene presence–absence as opposed to tree topologies.

5. Numerous caveats must be considered when interpreting patterns of lateral gene transfers

There are many caveats to be considered when interpreting ancient gene transfer events. Insights on both EGT and LGT are dependent on taxon sampling, which is uneven in our dataset in terms of the availability and quality of data from diverse lineages. The impact of taxon sampling on inferences about ancient LGT can be seen in the changing narrative about a single gene transfer of a tyrosyl-tRNA synthetase gene, which was originally argued to be a synapomorphy for Opisthokonta based on available data [61]. Reanalysis with data from additional microbial eukaryotes revealed that this gene was also present in Amoebozoa [40]. In our expanded taxon sampling, we find this gene in multiple lineages in Amoebozoa plus the parasite Blastocystis homoni (Sr_st), the orphan lineage *Palpitomonas bilix* (EE_is_Pbil) and two Rhizarian species (Sr_rh; electronic supplementary material, figure S1). While additional work is needed to rule out that these additional sparsely distributed taxa are not spurious data (i.e. contamination), our changing understanding of the phylogenetic distribution of the tyrosyl-tRNA synthetase gene highlights the impact of taxon sampling on inferences of ancient gene transfer events.

We also checked to see whether we find a pattern of interdomain LGT from Chlamydiae to Archaeplastida and other photosynthetic eukaryotes, which was observed in several previous analyses [33,34,62,63]. Only 10 genes of our 1138 matched the criteria of being present in three or fewer major eukaryotic clades with a single species or monophyletic clade of Chlamydiae as sister taxa (electronic supplementary material, table S2). Of these, only one (OG5_146631, an FAD-dependent oxidoreductase family protein) is found exclusively in photosynthetic eukaryotes. We also looked at the gene trees created by our pipeline for the 38 proteins reported as possible cases of transfer from Chlaymdiae to photosynthetic eukaryotes, and found that only half could be argued to be consistent with this transfer hypothesis given our taxonomic sampling (electronic supplementary material, table S3). Importantly, only two of the 38 genes reported by Becker et al. [34] match the more conservative criteria employed in our analyses as eukaryotes are not monophyletic in the remaining 36 trees. For our analyses of interdomain LGTs, we rejected trees with non-monophyletic eukaryotes as we do not believe we can distinguish between multiple interdomain LGTs and poorly resolved phylogenies without more in depth phylogenetic analyses.

Gene loss is clearly a major force in genome evolution [13,42], so interpreting the ancient LGT events must be done with caution particularly given the variation in genome sizes among the eukaryotes sampled for this study. Based on inferences on patterns of intron loss in eukaryotes and on genome complexity in the LECA, Wolf & Koonin [42] argue that gene loss has dominated the evolution of eukaryotic genomes, with intervening periods of 'complexification' that may include pulses of LGT/EGT. Because of the importance of gene loss we recognize that some of our examples of recent interdomain LGT may instead be more ancient gene transfer events that were then lost in major clades of eukaryotes. At the same time, we anticipate that even more recent events will be found as taxon sampling

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expands, particularly in poorly sampled territories like much of the Excavata and Rhizaria [43].

6. Conclusion

We identify 1138 genes that meet our criteria of possible interdomain LGTs as they are found in prokaryotes plus three or fewer major clades of eukaryotes. Analyses of the patterns among these genes reveals evidence of recent interdomain LGT events between prokaryotes and eukaryotes (table 3 and figure 2) and no compelling evidence of retained ancient LGTs (i.e. those that occurred in eukaryotic ancestors prior to the divergence of major clades). In contrast, we do detect numerous examples of EGTs involving multiple lineages of photosynthetic eukaryotes (figure 2 and figure 3), which validates our phylogenomic approach to detecting interdomain gene transfer events as the impact of EGTs has been established using other approaches [21–23,29]. With the exception of the EGTs, the data presented here are consistent with a model whereby gene loss is a dominant force in the evolution of eukaryotic genomes [42] as our analyses indicate that most interdomain LGTs have been lost over evolutionary time.

Note added in proof

The findings reported here are generally concordant with those from analyses of approximately 100 000 genes in a more limited number of eukaryotes plus prokaryotes (Ku *et al.* In press. Nature).

Data accessibility. Alignments and trees for the 1138 genes are available in the Dryad Digital Repository (http://dx.doi.org/10.5061/dryad. 2bj36).

Competing interests. We declare we have no competing interests.

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