A New Genetic Locus in *Sinorhizobium meliloti* is Involved in Stachydrine Utilization

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A New Genetic Locus in *Sinorhizobium meliloti* Is Involved in Stachydrine Utilization

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Received 24 February 1998/Accepted 12 July 1998

Stachydrine, a betaine released by germinating alfalfa seeds, functions as an inducer of nodulation genes, a catabolite, and an osmoprotectant in *Sinorhizobium meliloti*. Two stachydrine-inducible genes were found in *S. meliloti* 1021 by mutation with a Tn5-*luxAB* promoter probe. Both mutant strains (S10 and S11) formed effective alfalfa root nodules, but neither grew on stachydrine as the sole carbon and nitrogen source. When grown in the absence or presence of salt stress, S10 and S11 took up [14C]stachydrine as well as wild-type cells did, but neither used stachydrine effectively as an osmoprotectant. In the absence of salt stress, both S10 and S11 took up less [14C]proline than wild-type cells did. S10 and S11 appeared to colonize alfalfa roots normally in single-strain tests, but when mixed with the wild-type strain, their rhizosphere counts were reduced more than 50% (*P* < 0.01) relative to the wild type. These results suggest that stachydrine catabolism contributes to root colonization. DNA sequence analysis identified the mutated locus in S11 as *putA*, and the *luxAB* fusion in that gene was induced by proline as well as stachydrine. DNA that restored the capacity of mutant S10 to catabolize stachydrine contained a new open reading frame, *stcD*. All data are consistent with the concept that *stcD* codes for an enzyme that produces proline by demethylation of N-methylproline, a degradation product of stachydrine.

Plants release a variety of compounds that affect soil microorganisms and contribute to structuring microbial communities in the rhizosphere. More than 400 different molecules identified in alfalfa (*Medicago sativa* L.) may eventually reach soil bacteria through exudation or decomposition (38). Some of these compounds serve as transcriptional regulators of genes in *Rhizobium meliloti* (29), which was recently reclassified as *Sinorhizobium meliloti* (8). Other compounds are cofactors for microbial enzymes (40), and a few function as osmoprotectants (33). Any plant-derived compound present in sufficient concentration probably can serve as an energy substrate for rhizosphere microbes (5). However, defining molecules that have selective or specific effects on bacterial growth in the alfalfa rhizosphere and identifying rhizobial genes that are involved in responding to these factors is essential to understanding how communities of microorganisms develop on plant roots.

Several genetic tools have been used to define molecular plant-microbe interactions in the rhizosphere. Mutagenesis experiments have located a number of bacterial genes affecting root colonization (23, 31). In addition, enrichment for bacterial transconjugants (3) or for DNA amplifications (30) has found genes that enhance rhizosphere growth. The importance of a specific molecule in the rhizosphere can also be assessed by transforming plants to overproduce the compound (17) or by measuring the competitiveness of bacterial mutants incapable of catabolizing that molecule (10, 24).

Stachydrine, also known as N,N-dimethylproline or proline betaine, is a quaternary ammonium derivative of proline that occurs widely in *Medicago* species (39) but not in other genera of the Leguminosae (48). Most stachydrine-accumulating plants are found in the Capparidaceae (caper) and Labiatae (mint) families (48). Stachydrine was identified as an inducer of *S. meliloti* nodulation (*nod*) genes in alfalfa seed rinses (37), and many *Medicago* species nodulated by *S. meliloti* deposit this compound on developing seeds, where it is available to soil bacteria during seed germination (39). Stachydrine is probably a general osmoprotectant in alfalfa, because it is found not only on dried seed coats and in cured hay (43) but also in water-stressed alfalfa root nodules (20). Thus, *S. meliloti* may be exposed to stachydrine both during colonization of the young alfalfa root and later as N2-fixing cells in the root nodules.

Because *S. meliloti* uses stachydrine as a carbon and nitrogen source (16) and as an osmoprotectant (14), this molecule may be ecologically important for root colonization by this microorganism. In the absence of osmotic stress, *S. meliloti* cells catabolize stachydrine by sequential demethylations to N-methylproline and proline, but osmotically stressed cells accumulate stachydrine while catabolism is strongly reduced (14). To examine the role of stachydrine in root colonization, we mutagenized *S. meliloti* with the transposon Tn5-*luxAB* (47), which generates both insertional mutations and transcriptional luciferase reporter gene fusions. Here we describe the identification of two stachydrine-inducible genes in *S. meliloti* and show that the corresponding Tn5-*luxAB* insertional mutants are impaired in the use of stachydrine as a catabolite and as an osmoprotectant. The data also suggest that both mutants are impaired in competition for colonization of the rhizosphere. One of the stachydrine-inducible genes is identified as *putA*, a gene that was recently shown to promote root coloni-
zation (19). The other stachydrine-inducible locus (stcD) has not been previously identified and is shown to encode a putative demethylase that converts N-methylproline to proline. Unlike other genes involved in stachydrine catabolism (15), stcD is not located on the symbiotic plasmid of *S. meliloti*.

### MATERIALS AND METHODS

#### Bacterial strains and plasmids

The strains and plasmids used in this study are listed in Table 1. *S. meliloti* and *Escherichia coli* were maintained, respectively, in TY medium (4) and Luria-Bertani LB medium (41). *S. meliloti* was grown in GTS (Bethesda Research Laboratories, Gaithersburg, Md.). *E. coli* was grown in TY medium (4) and Luria-Bertani LB medium (41).

#### Generation of mutants and reporter gene insertions

Plasmid pRL1063a, which contains the transposable promoter probe Tn5-1063 with a promoterless luxAB reporter and the oriV of *E. coli* (47), was used to generate 5,000 insertional mutants of *S. meliloti* 1021 (27). The mutants were screened for stachydrine-mediated induction of lux expression after being grown for 3 days on Rec-5 filters (Nuclease Corp., Pleasanton, Calif.) on GTS agar medium. The filters were moved to fresh GTS agar with or without 100 μM stachydrine for 6 h and then exposed to the luciferase reporter substrate n-decanal (Sigma Co., St. Louis, Mo.). Luminescence was recorded for 4 h on sensitized (25) XAR X-ray film (Eastman Kodak, Rochester, N.Y.). Mutants showing stachydrine-dependent bioluminescence were retested twice and compared with a positive-control mutant in which *luxAB* was expressed constitutively. The stachydrine used in these studies was synthesized by standard procedures (35). Responses of mutant strains S10 and S11 to different concentrations of stachydrine and L-proline in appropriate media were tested. Additional growth tests were conducted in VM liquid and VM agar lacking mannitol and potassium nitrate. Complete VM agar was used as a positive control. Additional growth tests were conducted in VM liquid and VM agar lacking mannitol and potassium nitrate. Complete VM agar was used as a positive control.

### TABLE 1. Bacterial strains and plasmids used in this study

<table>
<thead>
<tr>
<th>Strain or plasmid</th>
<th>Relevant trait(s)</th>
<th>Source or reference</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>E. coli</em></td>
<td></td>
<td></td>
</tr>
<tr>
<td>HB101</td>
<td>pro leu thi lacY Str&lt;sup&gt;+&lt;/sup&gt;, endo recA r&lt;sup&gt;+&lt;/sup&gt;, m&lt;sup&gt;+&lt;/sup&gt;</td>
<td>9</td>
</tr>
<tr>
<td>YMC11</td>
<td>pro thi endA hsr Δ(glnA-glnD)</td>
<td>2</td>
</tr>
<tr>
<td><em>S. meliloti</em></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Rm1021</td>
<td>2011, Sm&lt;sup&gt;+&lt;/sup&gt;</td>
<td>32</td>
</tr>
<tr>
<td>Rm1021-S10 (S10)</td>
<td>1021::Tn5pRL1063a</td>
<td>This work</td>
</tr>
<tr>
<td>Rm1021-S11 (S11)</td>
<td>1021::Tn5pRL1063a</td>
<td>This work</td>
</tr>
<tr>
<td><strong>Plasmids</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>pBSK+</td>
<td>Multicyclic cloning vector</td>
<td>Stratagene</td>
</tr>
<tr>
<td>pRL1063a</td>
<td>Tn5-luxAB, Sm&lt;sup&gt;-&lt;/sup&gt; Km&lt;sup&gt;+&lt;/sup&gt; OriV</td>
<td>47</td>
</tr>
<tr>
<td>pLAFR1</td>
<td>Tc&lt;sup&gt;+&lt;/sup&gt;, Cos Tra&lt;sup&gt;-&lt;/sup&gt;, Mob&lt;sup&gt;-&lt;/sup&gt;, IncP</td>
<td>13</td>
</tr>
<tr>
<td>pRK2013</td>
<td>pRK212.2 conjugation helper plasmid</td>
<td>11</td>
</tr>
<tr>
<td>pSIF10</td>
<td>Circularized 10-kb EcoRI fragment of Rm1021-S10::Tn5-1063</td>
<td>This work</td>
</tr>
<tr>
<td>pSIF11</td>
<td>Circularized 12-kb EcoRI fragment of Rm1021-S11::Tn5-1063</td>
<td>This work</td>
</tr>
<tr>
<td>pLES1</td>
<td>pLAFR1 cosmid of Rm1021 containing stcD</td>
<td>This work</td>
</tr>
<tr>
<td>pLEB1</td>
<td>6.3-kb EcoRI-BamHI fragment carrying stcD in pBSK+</td>
<td>This work</td>
</tr>
<tr>
<td>pLEB55</td>
<td>6.8-kb EcoRI fragment carrying nodD2 in pBSK+</td>
<td>16</td>
</tr>
<tr>
<td>pLBR56</td>
<td>6.5-kb EcoRI fragment near nodD2 in pBSK+</td>
<td>16</td>
</tr>
</tbody>
</table>

---

**Strain or plasmid**  | **Relevant trait(s)**  | **Source or reference**
---|---|---
**E. coli**  |  |  |
HB101  | pro leu thi lacY Str<sup>+</sup>, endo recA r<sup>+</sup>, m<sup>+</sup>  | 9 |
YMC11  | pro thi endA hsr Δ(glnA-glnD)  | 2 |
**S. meliloti**  |  |  |
Rm1021  | 2011, Sm<sup>+</sup>  | 32 |
Rm1021-S10 (S10)  | 1021::Tn5pRL1063a  | This work |
Rm1021-S11 (S11)  | 1021::Tn5pRL1063a  | This work |
**Plasmids**  |  |  |
pBSK+  | Multicyclic cloning vector  | Stratagene |
pRL1063a  | Tn5-luxAB, Sm<sup>-</sup> Km<sup>+</sup> OriV  | 47 |
pLAFR1  | Tc<sup>+</sup>, Cos Tra<sup>-</sup>, Mob<sup>-</sup>, IncP  | 13 |
pRK2013  | pRK212.2 conjugation helper plasmid  | 11 |
pSIF10  | Circularized 10-kb EcoRI fragment of Rm1021-S10::Tn5-1063  | This work |
pSIF11  | Circularized 12-kb EcoRI fragment of Rm1021-S11::Tn5-1063  | This work |
pLES1  | pLAFR1 cosmid of Rm1021 containing stcD  | This work |
pLEB1  | 6.3-kb EcoRI-BamHI fragment carrying stcD in pBSK+  | This work |
pLEB55  | 6.8-kb EcoRI fragment carrying nodD2 in pBSK+  | 16 |
pLBR56  | 6.5-kb EcoRI fragment near nodD2 in pBSK+  | 16 |

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**Strain or plasmid**  | **Relevant trait(s)**  | **Source or reference**
---|---|---
**E. coli**  |  |  |
HB101  | pro leu thi lacY Str<sup>+</sup>, endo recA r<sup>+</sup>, m<sup>+</sup>  | 9 |
YMC11  | pro thi endA hsr Δ(glnA-glnD)  | 2 |
**S. meliloti**  |  |  |
Rm1021  | 2011, Sm<sup>+</sup>  | 32 |
Rm1021-S10 (S10)  | 1021::Tn5pRL1063a  | This work |
Rm1021-S11 (S11)  | 1021::Tn5pRL1063a  | This work |
**Plasmids**  |  |  |
pBSK+  | Multicyclic cloning vector  | Stratagene |
pRL1063a  | Tn5-luxAB, Sm<sup>-</sup> Km<sup>+</sup> OriV  | 47 |
pLAFR1  | Tc<sup>+</sup>, Cos Tra<sup>-</sup>, Mob<sup>-</sup>, IncP  | 13 |
pRK2013  | pRK212.2 conjugation helper plasmid  | 11 |
pSIF10  | Circularized 10-kb EcoRI fragment of Rm1021-S10::Tn5-1063  | This work |
pSIF11  | Circularized 12-kb EcoRI fragment of Rm1021-S11::Tn5-1063  | This work |
pLES1  | pLAFR1 cosmid of Rm1021 containing stcD  | This work |
pLEB1  | 6.3-kb EcoRI-BamHI fragment carrying stcD in pBSK+  | This work |
pLEB55  | 6.8-kb EcoRI fragment carrying nodD2 in pBSK+  | 16 |
pLBR56  | 6.5-kb EcoRI fragment near nodD2 in pBSK+  | 16 |
RESULTS

Rhizosphere colonization experiments. Root colonization tests were conducted with Moapa 69 alfalfa plants in axenic vermiculite by recovering bacteria from roots of individual plants as described previously (45), except that sterilized seeds were planted directly to minimize stachydrine loss and inocula were grown in TY medium. Each seed was inoculated with 300 to 500 bacterial cells at the time of planting. CFU were determined by plating cells on TY agar with Congo red (10 μg/ml) and appropriate antibiotics. The roots were rinsed in sterile water, and then more than 95% of the viable bacteria were removed from the roots by vortexing and sonication (45). Treatments consisted of at least three replicate jars, each containing five plants. Data obtained with the wild-type and mutant strains (CFU/root) were transformed and compared by using an unpaired t test for single-strain trials and a paired t test for competitive colonization trials. All experiments were repeated at least once.

Nucleotide sequence accession number. Sequence data for the 2,898-bp DNA fragment containing ORF I, here termed stcD, were submitted to the GenBank (NCBI) database as accession no. AF016307.

In vitro phenotypes. Twelve mutants containing stachydrine-inducible genes were isolated from \textit{S. meliloti} 1021. Each mutant showed a single Tn\textsubscript{5} insertion in hybridization tests after digesting total DNA with \textit{EcoRI} or \textit{Clai}, which cut outside Tn5-1063 (reference 6 and data not shown). All 12 mutants formed effective nodules on Moapa 69 alfalfa plants; 9 mutants had an impaired capacity to use stachydrine as an osmoprotectant in LAS medium containing 500 mM NaCl. Only strains S10 and S11 failed to grow on stachydrine as the sole carbon and nitrogen source, and they were selected for further studies. Maximum induction of the \textit{luxAB} reporter in strain S10 (Fig. 1A) was produced with a lower concentration of stachydrine (10 μM) than in S11 (100 μM) (Fig. 1B). Mutant S11, but not S10, also expressed luciferase in response to proline (Table 2).

Strains S10 and S11 differed in their capacity to grow on catabolic products of stachydrine (Table 3). S10 grew on proline but not on N-methylproline, while S11 grew on neither compound. The proline uptake capacity in strains S10 and S11, as measured with 10 μM \textit{[14C]proline}, was impaired by 41 and 78%, respectively, relative to that in parent cells (Table 2). Tests for growth under salt stress showed that strains S10 (Fig. 2) and S11 (data not shown) behaved similarly. In the absence of salt stress, both strains grew as well as the parent strain, Rm1021. Stachydrine markedly protected parent cells from inhibition by 500 mM NaCl but had no such positive effect on S10 cells (Fig. 2B). Stachydrine (1 mM) protected S11 cells slightly from the inhibitory effects of 500 mM NaCl but not to the extent that it protected the parent cells (data not shown). Further tests showed that the mutations had no effect on \textit{[14C]stachydrine} uptake and indicated that both mutant and parent cells had an increased total stachydrine uptake approximately 10-fold when grown in the presence of 300 mM NaCl (Table 4). Analysis of cells exposed to \textit{[14C]stachydrine} for 18 h showed that both the parent strain Rm1021 and the mutant strain S11 transformed the \textit{14C} into ethanol-insoluble materials whereas the capacity of mutant S10 to do this was significantly impaired (Table 4).

Rhizosphere phenotypes. Mutant strains S10 and S11 colonized roots as well as parent Rm1021 cells when they were inoculated separately on seeds (data not shown). In all cases, an inoculum of several hundred cells increased to nearly 10\textsuperscript{6} CFU/root by day 6. However, when the mutant strains were co-inoculated with the parent strain in a competition experiment, both mutants showed impaired root colonization (Fig. 3). Strain S10 colonized roots with significantly fewer cells (\(P \leq 0.05\)).

<table>
<thead>
<tr>
<th>Strain</th>
<th>Rate (nmol of proline/min/mg)(\text{a})</th>
<th>Decrease (%)</th>
<th>\textit{luxAB} induction (RLU/OD unit/min)(\text{b})</th>
</tr>
</thead>
<tbody>
<tr>
<td>Rm1021</td>
<td>27 ± 1</td>
<td>0</td>
<td>3,070 ± 584</td>
</tr>
<tr>
<td>Rm1021-S10</td>
<td>16 ± 4</td>
<td>41</td>
<td>1,390 ± 446</td>
</tr>
<tr>
<td>Rm1021-S11</td>
<td>6 ± 1</td>
<td>78</td>
<td>2,490 ± 40</td>
</tr>
</tbody>
</table>

\(\text{a}\) Uptake measured with 10 μM \textit{[14C]proline}.

\(\text{b}\) Maximum response observed during a 6-h induction test with 10 or 100 μM proline for Rm1021-S10 and Rm1021-S11, respectively.

NA, not applicable.
than did Rm1021 on days 2, 4, and 6 (Fig. 3A). A 1.01 ratio of S10 to Rm1021 cells was measured on day 0, compared with significantly lower values (P < 0.01) of 0.26, 0.45, and 0.34 on days 2, 4, and 6, respectively (Fig. 3A). The 1.16 ratio of S11 to Rm1021 on day 0 declined to significantly lower values (P < 0.01) of 0.48 and 0.43 on days 4 and 6, respectively (Fig. 3B). Tests with other stachydrine-inducible Tn5-luxAB mutants from this study showed that some, but not all, were impaired in competitive colonization tests with the parent, Rm1021 (data not shown).

**Molecular analyses.** DNA hybridization analyses located the Tn5 insertions of strains S10 and S11 in 10-kb and 12-kb EcoRI restriction fragments, respectively (Table 1). DNA sequence data from pSIF11 showed that the Tn5-luxAB insertion in strain S11 resided in a gene having nearly complete DNA nucleotide identity to the proline dehydrogenase gene (putA) of S. meliloti GR4 (19). Of 842 nucleotides, 813 (96.5%) totally matched those previously reported for strain GR4. The deduced amino acid homology was 96%.

The PCR-generated hybridization probe corresponding to the flanking regions of the Tn5-tagged locus in strain S10 identified cosmid pLES1 as containing the corresponding wild-type Rm1021 DNA. Transconjugants of strain S10 containing cosmid pLES1 grew on stachydrine as the sole carbon and nitrogen source (data not shown), and DNA hybridization analysis of the complemented mutant showed that both the original mutated gene and the wild-type locus were present (Fig. 4). Electrophoretic analyses of plasmid versus total genomic DNA from the wild-type strain Rm1021 showed that the probe for the mutated locus in strain S10 hybridized to DNA which was physically distinct from the symbiotic plasmid that hybridized to the nodC-specific probe (data not shown). Other tests showed the probe produced from the mutated locus hybridized with neither pLBR55 nor pLBR56, both of which contain stachydrine catabolism genes from the nodC-containing symbiotic plasmid of S. meliloti (16) (data not shown).

Nucleotide sequence analysis of a 2,898-bp DNA fragment in pLEB1 identified ORF I, which in mutant S10 contained the 0.01) than did Rm1021 on days 2, 4, and 6 (Fig. 3A). A 1.01 ratio of S10 to Rm1021 cells was measured on day 0, compared with significantly lower values (P < 0.01) of 0.26, 0.45, and 0.34 on days 2, 4, and 6, respectively (Fig. 3A). The 1.16 ratio of S11 to Rm1021 on day 0 declined to significantly lower values (P < 0.01) of 0.48 and 0.43 on days 4 and 6, respectively (Fig. 3B). Tests with other stachydrine-inducible Tn5-luxAB mutants from this study showed that some, but not all, were impaired in competitive colonization tests with the parent, Rm1021 (data not shown).

**Table 4.** Stachydrine uptake and catabolism by S. meliloti

<table>
<thead>
<tr>
<th>Strain</th>
<th>NaCl addeda</th>
<th>Stachydrine uptakeb after:</th>
<th>% of 14C incorporatedc</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>10 min</td>
<td>18 h</td>
<td>Soluble: Insoluble</td>
</tr>
<tr>
<td>Rm1021</td>
<td>-</td>
<td>2 2</td>
<td>54:46</td>
</tr>
<tr>
<td></td>
<td>+</td>
<td>18 30</td>
<td>63:37</td>
</tr>
<tr>
<td>Rm1021-S10</td>
<td>-</td>
<td>1 3</td>
<td>87:13</td>
</tr>
<tr>
<td></td>
<td>+</td>
<td>16 49</td>
<td>95:5</td>
</tr>
<tr>
<td>Rm1021-S11</td>
<td>-</td>
<td>1 1</td>
<td>61:39</td>
</tr>
<tr>
<td></td>
<td>+</td>
<td>20 25</td>
<td>70:30</td>
</tr>
</tbody>
</table>

a NaCl added to LAS medium at 300 mM 24 h prior to supplementation with [14C]stachydrine.
b Uptake measured as nanomoles per milligram of protein per minute at 10 min and 18 h.
c Ethanol-soluble and -insoluble cell fractions determined in the 18-h uptake sample.
Bacterium a, characterized as enzyme reported from Thermoanaerobium. Rm1021-S10 (lane 2), and transconjugant Rm1021-S10(pLES1) (lane 3) was the catabolism transconjugant. Total genomic DNA from Rm1021, markers are shown in lane M.

Alignment analysis (1) of ORF I showed highly significant (with the production of a protein containing 580 amino acids. Tn insertion (Fig. 5). Data from ORF I are consistent with the production of a protein containing 580 amino acids. Alignment analysis (1) of ORF I showed highly significant (P < 4 × 10^-60) amino acid homology to NADH-dependent oxidases reported from Thermoanaerobium (39%) (28) and Eubacterium (35%) (12), which are assigned to a family of flavoproteins characterized as α/β-barrel oxidoreductases.

**DISCUSSION**

Of 12 stachydrine-inducible Tn5-luxAB insertions found among 5,000 S. meliloti mutants, two could not use stachydrine as the sole carbon and nitrogen source. These mutant strains, S10 and S11, colonized alfalfa roots normally in single-strain tests but showed reduced competitiveness in colonization tests with the parent, Rm1021 (Fig. 3). Growth, uptake, and DNA sequence data established that S11 was mutated in the known S. meliloti putA gene (19), and evidence here is consistent with the conclusion that the mutated ORF in S10, stcD, is involved in the second demethylation of stachydrine (i.e., demethylation of N-methylproline to form proline).

Data obtained with mutant strain S11 (Fig. 3B) confirm the known contribution of putA to competitive alfalfa root colonization (19) and extend our understanding by showing that exogenous stachydrine induces that gene. The impairment of proline uptake in this mutant (Table 2) suggests that PutA in S. meliloti plays a regulatory role, as has been shown for this multifunctional protein in enteric bacteria (34) but not in Bradyrhizobium (44). Whether the putA gene in S. meliloti is induced directly by stachydrine or indirectly by degradation of stachydrine to proline, as suggested by proline effects on luxAB expression (Table 2), was not established by these experiments. However, the ecological significance of putA presumably is proportional to the sum total of stachydrine and proline released by the plant. Because high levels of stachydrine are released from germinating Medicago seeds (39), significant amounts of proline may be produced by catabolism. Preliminary results have shown that [14C]stachydrine is effectively catabolized to proline by young alfalfa plants (26) and that S. meliloti may therefore be exposed to N-methylproline as well as to stachydrine.

Data presented here indicate that the mutated locus in S10 is genetically and phenotypically distinct from stachydrine catabolism genes identified previously in S. meliloti (16). First, the DNA-DNA hybridization analysis of plasmid and total DNA indicates that DNA containing the mutated locus in S10 is not on the symbiotic plasmid and may be on the chromosome or another plasmid. Second, symbiotic plasmid fragments containing known stachydrine catabolism genes (16) did not hybridize with the mutated locus in S10. Finally, while previously reported mutants grew on both N-methylproline and proline (16), S10 failed to grow on N-methylproline (Table 2). These data and the identification here of a previously unreported ORF containing the Tn5 insertion in S10 indicate that a novel gene, referred to here as stcD, is mutated in S10.

The function of stcD is not completely established by this study, but three lines of evidence are consistent with the concept that stcD codes for a protein required for the demethylation of N-methylproline. First, the physiological data indicate that strain S10 is impaired in the second demethylation of stachydrine because it grows on proline but not on N-methylproline as the sole carbon and nitrogen source (Table 3). Second, while S10 is capable of taking up stachydrine at levels similar to the parent, it does not catabolize the stachydrine (Table 4). In S10 cells, most [14C] remains as ethanol-soluble material and is not incorporated into the ethanol-insoluble cellular fraction. This result contrasts with results for both the parent strain and the S11 putA mutant, in which a large fraction of the [14C] was incorporated into cellular material. The mutation in putA would prevent the catabolism of proline but not the direct use of proline for protein synthesis. Third, the Tn5 insertion in S10 (Fig. 5) interrupted an ORF (GenBank accession no. AF016307) with significant deduced amino acid homology to genes encoding α/β-barrel oxidoreductase face.
voproteins. In *Pseudomonas*, at least two oxidases, 4-methoxy-benzoate demethylase and vanillate demethylase, function as demethylases (18). Although previous work identified genes involved in the conversion of stachydrine to N-methylproline (16), a locus responsible for the demethylation of N-methylproline has not been reported. Complementation of S10 by a cosmid containing the sequenced ORF (Fig. 4) was also consistent with this interpretation.

Root colonization results with the putA mutant, S11, in this study differed markedly from those reported by others (19). The much smaller effect of the mutation on colonization observed here may have resulted from the test we used. Our assay measured the capacity of a small initial inoculum to colonize roots growing in vermiculite, while the previous work (19) used roots exposed to a large initial inoculum in a hyponic medium. Under our conditions, both the Rm1021 parent and the S11 mutant grew significantly but the mutant cells achieved a final population on the root approximately half the size of that produced by the parent cells. Jimenez-Zurdo et al. (19) saw no growth of the putA mutant after inoculation while the number of wild-type cells increased 10-fold in 48 h. These results suggest that different factors were affecting bacterial growth and survival in the two studies.

These results offer additional evidence suggesting that stachydrine catabolism contributes to rhizobial colonization of alfalfa seedling roots. *S. meliloti* mutants incapable of growing on stachydrine as the sole carbon and nitrogen source are less efficient at forming root nodules on two *Medicago* species (16) and less competitive against isogenic wild-type strains (Fig. 3) (19). It is not surprising that utilization of a unique carbon resource such as stachydrine facilitates root colonization (10, 24). The moderate benefits measured in this study (Fig. 3), however, suggest either that stachydrine is not a major controlling factor in root colonization under the conditions tested or that higher levels of stachydrine are required for *S. meliloti* to benefit fully from the capacity to metabolize this compound. Experiments in nonsterile soil might help to assess the ecological significance of this trait in more complex microbial communities, and the use of plants modified to produce more stachydrine might overcome problems associated with limiting amounts of this compound.

**ACKNOWLEDGMENTS**

This work was supported by grants from the U.S. Department of Agriculture (NRICGP91-37305) and the U.S. National Science Foundation (IBN-92-18567). The Ministère de L’Enseignement Supérieur et de la Recherche (DREIF, France) is thanked for financial support for cooperative research.

We thank M. Thomasow for use of the luminometer and D. Tepfer for supplying pLBR55 and pLBR56.

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